

FILE 'MEDLINE, EMBASE, BIOSIS, BIOTECHDS, SCISEARCH, HCAPLUS, NTIS, LIFESCI' ENTERED AT 14:17:57 ON 06 FEB 2001

L1 106607 S REVERSE(W)TRANSCRIPTASE?  
L2 25 S ASLV AND L1  
L3 5 S L2 AND (PURIF? OR ISOLAT?)  
L4 3 DUP REM L3 (2 DUPLICATES REMOVED)  
L5 0 S UNITS AND L4  
L6 1 S L4 AND (NANOGRAM? OR MILLIGRAM? OR NG OR MG)  
L7 1 S UNTITS(W)MG  
L8 10647 S UNITS(W)MG  
L9 24 S L1 AND L8  
L10 0 S L9 AND ASLV  
L11 11 DUP REM L9 (13 DUPLICATES REMOVED)  
L12 190 S AMV(W)RT  
L13 0 S L12 AND L8  
L14 4 S AVAIN (W) RETROVIRUS  
L15 0 S L14 AND L8  
L16 1850 S AVIAN (W) RETROVIRUS  
L17 195 S L16 AND L1  
L18 0 S L17 AND L8  
L19 110 S L17 AND ACTIV?  
L20 1 S L19 AND UNIT?  
L21 0 S L2 AND (SPECIFIC(W)ACTIV?)  
L22 289 S L1 AND (AVIAN(W)SARCOMA )  
L23 0 S L22 AND L8  
L24 0 S "AVAIN SARCOMA LEUKOSIS VIRUS?"  
L25 147 S "AVIAN SARCOMA LEUKOSIS VIRUS"  
L26 26 S L25 AND (ISOLAT? OR PURIF?)  
L27 0 S L26 AND L8  
L28 0 S L26 AND (SPECIFIC(A)ACTIV?)  
L29 36 S L25 AND L1  
L30 30 S L29 AND ACTIV?  
L31 0 S L30 AND UNIT?  
L32 11 DUP REM L30 (19 DUPLICATES REMOVED)  
L33 1 S L29 AND (MG OR NG OR NANOGRAM? OR MILLIGRAM?)  
L34 1 S L30 AND (MG OR NG OR NANOGRAM? OR MILLIGRAM?)  
L35 8 S L30 AND SUBUNIT?  
L36 20 S RT(A) (UNIT)  
L37 46 S RT(A) (UNIT?)  
L38 0 S L30 AND L37  
L39 0 S L11 AND L25  
L40 286 S L25 OR ASLV  
L41 0 S L40 AND L8  
L42 5 S L40 AND HOMOGENE?  
L43 1 DUP REM L42 (4 DUPLICATES REMOVED)  
L44 0 S L43 AND L8  
L45 0 S L34 AND (SPECIFIC(A)ACTIV?)  
L46 0 S L43 AND (SPECIFIC(A)ACTIV?)

DN 88230595  
 TI Properties of **avian sarcoma-leukosis virus** pp32-related pol-endonucleases produced in *Escherichia coli*.  
 AU Terry R; Soltis D A; Katzman M; Cobrinik D; Leis J; Skalka A M  
 CS Roche Research Center, Hoffmann-La Roche Inc., Nutley, New Jersey 07110.  
 NC CA38046 (NCI)  
 T32-GM-07250 (NIGMS)  
 CA-06927 (NCI)  
 SO JOURNAL OF VIROLOGY, (1988 Jul) 62 (7) 2358-65.  
 Journal code: KCV. ISSN: 0022-538X.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals; Cancer Journals  
 EM 198809  
 AB The gag-pol precursor protein of the **avian sarcoma-leukosis virus** is processed into three known pol-encoded mature polypeptides; the 95- and 63-kilodalton (kDa) beta and alpha subunits, respectively, of reverse transcriptase and the 32-kDa pp32 protein. The pp32 protein possesses DNA endonuclease activity and is produced from the precursor by two proteolytic cleavage events, one of which removes 4.1 kDa of protein from the C terminus. A 36-kDa protein (p36pol) which retains this C-terminal segment is detectable in small quantities in virions. We have constructed *Escherichia coli* plasmid clones that express the C-terminal domains of pol corresponding to pp32 and p36. These proteins have been purified by column chromatographic methods to near **homogeneity**. No significant differences could be detected in the enzymatic properties of the bacterially produced p32pol and p36pol proteins. Both possess DNA endonuclease activity and, like the pp32 protein isolated from virions, can cleave near the junction of two tandem **avian sarcoma-leukosis virus** long terminal repeats in double-stranded supercoiled DNA substrates. In the presence of Mg<sup>2+</sup>, both p32pol and viral pp32 cleave either strand of DNA 2 nucleotides 5' to the junction.  
 CT Check Tags: Support, Non-U.S. Gov't; Support, U.S. Gov't, P.H.S.  
 Amino Acid Sequence  
 Base Sequence  
 \*DNA-Directed DNA Polymerase: GE, genetics  
 DNA-Directed DNA Polymerase: ME, metabolism  
 DNA, Superhelical: ME, metabolism  
 DNA, Viral: ME, metabolism  
 \*Endodeoxyribonucleases: GE, genetics  
 Endodeoxyribonucleases: ME, metabolism  
*Escherichia coli*: GE, genetics  
 Molecular Sequence Data  
 Protein Processing, Post-Translational  
 \*Recombinant Proteins: GE, genetics  
 Recombinant Proteins: ME, metabolism  
 \*Retroviridae Proteins: GE, genetics  
 Retroviridae Proteins: ME, metabolism  
 \*Sarcoma Viruses, Avian: EN, enzymology  
 Sarcoma Viruses, Avian: GE, genetics  
 CN EC 2.7.7.7 (DNA-Directed DNA Polymerase); EC 3.1.- (Endodeoxyribonucleases); 0 (avian retrovirus proteins); 0 (DNA, Superhelical); 0 (DNA, Viral); 0 (Recombinant Proteins); 0 (Retroviridae Proteins)

L11 ANSWER 8 OF 11 HCAPLUS COPYRIGHT 2001 ACS

ACCESSION NUMBER: 1982:81758 HCAPLUS

DOCUMENT NUMBER: 96:81758

TITLE: Purification of **reverse transcriptase** from avian myeloblastosis virus (AMV)

AUTHOR(S): Qi, Defang; Pan, Tiecheng; Feng, Zongming; Qian, Bin;

Yu, Weiyuan  
CORPORATE SOURCE: Shanghai Inst. Biochem., Acad. Sin., Shanghai, Peop.

Rep. China  
SOURCE: Shengwu Huaxue Yu Shengwu Wuli Xuebao (1981), 13(3),

275-81

CODEN: SHWPAU; ISSN: 0582-9879

DOCUMENT TYPE: Journal

LANGUAGE: Chinese

AB **Reverse transcriptase** was purified .apprx.600-fold from AMV. The method employed affinity chromatog. on a column of covalently linked heparin-agarose followed by concn. of the enzyme by dialysis against 50% glycerol in phosphate buffer. This procedure could be completed within 1.5 days. The purified **reverse transcriptase** showed sp. activity of 12,000 **units/mg** protein and enzyme yield of 30,000 units/g AMV, and was free of detectable DNase and RNase. Conditions affecting the enzyme activity, e.g. metal ions, template specificity, and the length of primers were also studied and discussed.

L11 ANSWER 1 OF 11 MEDLINE

DUPLICATE 1

ACCESSION NUMBER: 1999292945 MEDLINE

DOCUMENT NUMBER: 99292945

TITLE: Barley coleoptile peroxidases. Purification, molecular cloning, and induction by pathogens.

AUTHOR: Kristensen B K; Bloch H; Rasmussen S K

CORPORATE SOURCE: Plant Biology and Biogeochemistry Department, PBK-301, Riso

National Laboratory, P.O. Box 49, DK-4000 Roskilde, Denmark.. brian.kristensen@risoe.dk

SOURCE: PLANT PHYSIOLOGY, (1999 Jun) 120 (2) 501-12.

Journal code: P98. ISSN: 0032-0889.

PUB. COUNTRY: United States

Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 199910

ENTRY WEEK: 19991002

AB A cDNA clone encoding the Prx7 peroxidase from barley (*Hordeum vulgare* L.)

predicted a 341-amino acid protein with a molecular weight of 36,515.

N- and C-terminal putative signal peptides were present, suggesting a vacuolar location of the peroxidase. Immunoblotting and **reverse-transcriptase** polymerase chain reaction showed that the Prx7 protein and mRNA accumulated abundantly in barley coleoptiles and in leaf

epidermis inoculated with powdery mildew fungus (*Blumeria graminis*). Two

isoperoxidases with isoelectric points of 9.3 and 7.3 (P9.3 and P7.3, respectively) were purified to homogeneity from barley coleoptiles.

P9.3 and P7.3 had Reinheitszahl values of 3.31 and 2.85 and specific activities

(with 2,2'-azino-di-[3-ethyl-benzothiazoline-6-sulfonic acid], pH 5.5, as

the substrate) of 11 and 79 **units/mg**, respectively.

N-terminal amino acid sequencing and matrix-assisted laser desorption/ionization time-of-flight mass-spectrometry peptide analysis

identified the P9.3 peroxidase activity as due to Prx7. Tissue and subcellular accumulation of Prx7 was studied using activity-stained isoelectric focusing gels and immunoblotting. The peroxidase activity due

to Prx7 accumulated in barley leaves 24 h after inoculation with powdery

mildew spores or by wounding of epidermal cells. Prx7 accumulated predominantly in the epidermis, apparently in the vacuole, and appeared to

be the only pathogen-induced vacuolar peroxidase expressed in barley tissues. The data presented here suggest that Prx7 is responsible for the

biosynthesis of antifungal compounds known as hordatines, which accumulate

abundantly in barley coleoptiles.

L11 ANSWER 2 OF 11 HCAPLUS COPYRIGHT 2001 ACS

ACCESSION NUMBER: 1999:98778 HCAPLUS

DOCUMENT NUMBER: 130:278414

TITLE: A study of the purification and enzyme activity of the

recombinant HIV - 1 **reverse transcriptase**

AUTHOR(S): Ke, Yuehai; Wang, Jiaquan; Zeng, Yi

CORPORATE SOURCE:  
Preventive

Institute of Virology, Chinese Academy of

SOURCE:

Medicine, Beijing, 100052, Peop. Rep. China

Bingdu Xuebao (1998), 14(4), 315-321

CODEN: BIXUEA; ISSN: 1000-8721

PUBLISHER:

Bingdu Xuebao Bianjibu

DOCUMENT TYPE:

Journal

LANGUAGE:

Chinese

AB HIV-1 **reverse transcriptase** was expressed in E. coli DH5.alpha. using an expression plasmid RP66, derived from pBV220. The engineered bacteria were cultivated in the fermentator and induced to produce the insol. inclusion body formation of the non-active RT.

After

inclusion body clarification, solubilization and refolding, the further

purifn. step was investigated by HIC (hydrophobic - interaction chromatog.) and anion - exchange chromatog. (Q-sepharose).

Meanwhile, the

whole purifn. steps were monitored by RP - HPLC. Finally the sol. active

RT (purifn. level about 95 %, specific enzyme activity 9.75.times.105 units/mg) was obtained.

L11 ANSWER 3 OF 11 MEDLINE

DUPLICATE 2

ACCESSION NUMBER: 94053388 MEDLINE

DOCUMENT NUMBER: 94053388

TITLE: Characterization of recombinant human neuron-specific enolase and its application to enzyme immunoassay.

AUTHOR: Aoki T; Kimura M; Kaneta M; Kazama H; Morikawa J;

Watabe H

CORPORATE SOURCE: Department of Biochemistry, Faculty of Pharmaceutical Sciences, Higashi-Nippon-Gakuen University, Hokkaido, Japan..

SOURCE: TUMOUR BIOLOGY, (1993) 14 (5) 261-70.

Journal code: TUB. ISSN: 1010-4283.

PUB. COUNTRY: Switzerland

Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals; Cancer Journals

ENTRY MONTH: 199402

AB Human gamma-enolase cDNA prepared by **reverse transcriptase**-polymerase chain reaction was cloned into the Escherichia coli expression vector pKK223-3. The resulting plasmid, pHTK503, expressed human gamma-enolase as a 46-kDa protein in

SDS-PAGE, and in the cells as the active gamma gamma form (designated as recombinant

human NSE; R-NSE). R-NSE was purified from E. coli by several chromatographic elutions. Finally, 6.0 mg of R-NSE from 8.1 g cells

was

purified with a specific activity of 86 units/mg protein. The structural properties of R-NSE were compared with the NSE purified from human brain tissue (B-NSE). The biochemical and enzymatic

characteristics were essentially the same, except for the isoelectric point (4.5 for B-NSE and 4.7 for R-NSE). In an NSE immunoassay system, R-NSE and standard NSE were almost equal in reactivity to the anti-NSE antibody. These results indicate that R-NSE can be used as standard

assay

material.

L11 ANSWER 4 OF 11 MEDLINE

DUPLICATE 3

ACCESSION NUMBER: 89340495 MEDLINE

DOCUMENT NUMBER: 89340495

TITLE: Co-expression of the subunits of the heterodimer of HIV-1

**reverse transcriptase** in Escherichia coli.

AUTHOR: Muller B; Restle T; Weiss S; Gautel M; Sczakiel G;  
Goody R

S

CORPORATE SOURCE: Abteilung Biophysik, Max-Planck Institut fur  
Medizinische  
Forschung, Heidelberg, Federal Republic of Germany.  
SOURCE: JOURNAL OF BIOLOGICAL CHEMISTRY, (1989 Aug 25) 264 (24)  
13975-8.  
Journal code: HIV. ISSN: 0021-9258.  
PUB. COUNTRY: United States  
Journal; Article; (JOURNAL ARTICLE)  
LANGUAGE: English  
FILE SEGMENT: Priority Journals; Cancer Journals  
ENTRY MONTH: 198911

AB Expression of the 66-kDa form of human immunodeficiency virus, type 1  
**reverse transcriptase** in Escherichia coli leads to  
isolation of small amounts of a 2 x 66-kDa homodimer and larger  
amounts of  
a heterodimer form of the enzyme in which the 66-kDa protein is  
complexed  
with its carboxyl-terminally truncated is complexed with its  
carboxyl-terminally truncated 51-kDa form. The latter arises via  
proteolysis by contaminating proteases. The heterodimer, which was  
characterized by gel filtration (apparent native molecular mass of  
120-130  
kDa), was the most active form of the enzyme (specific activity, 5000  
**units/mg**, cf. less than 2000 for the 66-kDa fragment).  
The 66-kDa fragment alone was shown to be only partially dimerized,  
with  
the activity residing mainly in the dimer fraction. Proteolysis of the  
66-kDa form resulting partially in the 51-kDa form led to an increase  
in  
**reverse transcriptase** activity. Expression of a  
truncated version of the protein containing the first 428 amino acids  
of  
the **reverse transcriptase** coding region led to a  
protein which had low but measurable **reverse**  
**transcriptase** activity (400-500 **units/mg**).  
Co-expression of the two proteins on a single plasmid led to  
expression in  
a 1:1 ratio. The 1:1 mixture behaved as a heterodimer, as shown by its  
chromatographic properties. It is likely that the mechanism for the  
production of heterodimers in vivo involves cleavage of 66-kDa  
monomers  
followed by rapid dimerization of the 51- and 66-kDa forms to give the  
heterodimeric form, which is stable toward further proteolysis.

L11 ANSWER 5 OF 11 LIFESCI COPYRIGHT 2001 CSA  
ACCESSION NUMBER: 89:42512 LIFESCI  
TITLE: Co-expression of the subunits of the heterodimer of  
HIV-1

**reverse transcriptase** in Escherichia coli

AUTHOR: Mueller, B.; Restle, T.; Weiss, S.; Gautel, M.;  
Sczakiel,

G.; Goody, R.S.

CORPORATE SOURCE: Abt. Biophys., Max-Planck Inst. Med. Forsch.,  
Janhnstr. 29,

6900 Heidelberg, FRG

SOURCE: J. BIOL. CHEM., (1989) vol. 264, no. 24, pp.  
13975-3978.

DOCUMENT TYPE: Journal

FILE SEGMENT: V; N

LANGUAGE: English

SUMMARY LANGUAGE: English

AB Expression of the 66-kDa form of human immunodeficiency virus, type 1  
**reverse transcriptase** in Escherichia coli leads to  
isolation of small amounts of a 2 x 66-kDa homodimer and larger  
amounts of  
a heterodimer form of the enzyme in which the 66-kDa protein is  
complexed  
with its carboxyl-terminally truncated 51-kDa form. The latter arises  
via  
proteolysis by contaminating proteases. The heterodimer, which was

characterized by gel filtration (apparent native molecular mass of 120-130 kDa), was the most active form of the enzyme (specific activity, 5000 units/mg, cf. <2000 for the 66-kDa fragment). The 66-kDa fragment alone was shown to be only partially dimerized, with the activity residing mainly in the dimer fraction.

L11 ANSWER 6 OF 11 HCAPLUS COPYRIGHT 2001 ACS

ACCESSION NUMBER: 1987:63587 HCAPLUS

DOCUMENT NUMBER: 106:63587

TITLE: Purification of **reverse transcriptase**

INVENTOR(S): Noda, Akihiro; Mukai, Hiroyuki; Ohayashi, Akira

PATENT ASSIGNEE(S): Takara Shuzo Co., Ltd., Japan

SOURCE: Jpn. Kokai Tokkyo Koho, 6 pp.

CODEN: JKXXAF

DOCUMENT TYPE: Patent

LANGUAGE: Japanese

FAMILY ACC. NUM. COUNT: 1

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
JP 61212284	A2	19860920	JP 1985-55167	19850319

AB During purifn. of **reverse transcriptase** from a RNA tumor virus, the ext. is subjected to gel filtration high-speed liq. chromatog. for purifn. Thus, an ext. from Rous-assocd. 2 virus was chromatographed on DEAE-Trisacryl M, subjected to high-speed liq. chromatog. using TSK Gel 3000 SWG, and treated with PEG 2000 for concn. to give purified **reverse transcriptase** with sp. activity of 74,400 units/mg.

L11 ANSWER 7 OF 11 HCAPLUS COPYRIGHT 2001 ACS

ACCESSION NUMBER: 1982:594918 HCAPLUS

DOCUMENT NUMBER: 97:194918

TITLE: Studies on the purification and the reverse transcription activity of E. coli DNA polymerase I

AUTHOR(S): Cai, Faxing; Yu, Xiyuan; Zhang, Guiqin; Cheng, Zhenqi;

Wei, Xiping; Zhao, Pi

CORPORATE SOURCE: Inst. Microbiol., Acad. Sin., Beijing, Peop. Rep. China

SOURCE: Shengwu Huaxue Yu Shengwu Wuli Jinzhan (1982), 44, 37-40

CODEN: SHYCD4

DOCUMENT TYPE: Journal

LANGUAGE: Chinese

AB DNA polymerase I (I) was isolated and purified from sonicated Escherichia

coli B cells by (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> pptn., and DEAE-cellulose 52, cellulose phosphate P-70, and hydroxylapatite column chromatog. in 8% yield.

The specific activity of the purified I was increased from 209 units/mg (following (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> pptn.) to 3000 units/mg after the hydroxylapatite column chromatog. The purified I prepn. did not show RNase activity. I activity was inhibited by 91%

in the presence of actinomycin D and was inhibited by 87% when dGTP, dCTP, or dTTP was omitted from the reaction mixt. The reverse transcription

activity of the I prepn. was detd. in a 0.1 mL Tris-HCl buffer (pH 7.4)

system using poly(A) as template, oligo(dT) as primer, and [3H]TTP as substrate and incubated at 23.degree. for 45 min. The reverse transcription system was activated by divalent ions in the order:

Mg<sup>2+</sup> >

Hg<sup>2+</sup> > Ca<sup>2+</sup> at optimal concn. of 1-2, 6, and 7 mM, resp., whereas

Ca<sup>2+</sup> was

inhibitory. The reverse transcription activity of I was decreased

when I

concn. in the system was >3 .mu.g; the decrease in the reverse transcription activity at high I concn. may have been due to the activation of the exonuclease activity of I. Thus, the E. coli I prepn.

cannot be used to replace avian myeloblastosis virus **reverse transcriptase**.

L11 ANSWER 8 OF 11 HCAPLUS COPYRIGHT 2001 ACS

ACCESSION NUMBER: 1982:81758 HCAPLUS

DOCUMENT NUMBER: 96:81758

TITLE: Purification of **reverse transcriptase** from avian myeloblastosis virus (AMV)

AUTHOR(S): Qi, Defang; Pan, Tiecheng; Feng, Zongming; Qian, Bin;

Yu, Weiyuan  
CORPORATE SOURCE: Shanghai Inst. Biochem., Acad. Sin., Shanghai, Peop.

Rep. China  
SOURCE: Shengwu Huaxue Yu Shengwu Wuli Xuebao (1981), 13(3),

275-81

CODEN: SHWPAU; ISSN: 0582-9879

DOCUMENT TYPE: Journal

LANGUAGE: Chinese

AB **Reverse transcriptase** was purified .apprx.600-fold from AMV. The method employed affinity chromatog. on a column of covalently linked heparin-agarose followed by concn. of the enzyme by dialysis against 50% glycerol in phosphate buffer. This procedure could

be completed within 1.5 days. The purified **reverse transcriptase** showed sp. activity of 12,000 **units/mg** protein and enzyme yield of 30,000 units/g AMV, and was free of detectable DNase and RNase. Conditions affecting the enzyme activity, e.g. metal ions, template specificity, and the length of primers were also

studied and discussed.

L11 ANSWER 9 OF 11 HCAPLUS COPYRIGHT 2001 ACS

ACCESSION NUMBER: 1979:134398 HCAPLUS

DOCUMENT NUMBER: 90:134398

TITLE: **Reverse transcriptase** from avian myeloblastosis virus

AUTHOR(S): Houts, G. Edwin; Miyagi, Masakazu; Ellis, Carmen; Beard, Dorothy; Beard, J. W.

CORPORATE SOURCE: Life Sci., Inc., St. Petersburg, Fla., USA

SOURCE: J. Virol. (1979), 29(2), 517-22

CODEN: JOVIAM; ISSN: 0022-538X

DOCUMENT TYPE: Journal

LANGUAGE: English

AB From lots of 20-30 g of avian myeloblastosis virus, RNA-dependent DNA polymerase was obtained in prepns. of purity >95% by using a 2-step column

chromatog. procedure employing DEAE- (DE 52) and CM-cellulose (CM 52). Yields of RNA-dependent DNA polymerase varied from .apprx.20,000 to 35,000

units/g of virus. Sp. activity of the enzyme was .apprx.35,000-60,000 **units/mg** of protein. Free of detectable RNase activity, the product exhibited a mol. wt. of .apprx.160,000, an isoelec. point of

6.5, and .apprx.2 mol of fatty acid/mol enzyme.

L11 ANSWER 10 OF 11 BIOSIS COPYRIGHT 2001 BIOSIS

ACCESSION NUMBER: 1978:184734 BIOSIS

DOCUMENT NUMBER: BA65:71734

TITLE: TERMINAL DEOXY RIBO NUCLEOTIDYL TRANSFERASE EC-2.7.7.31 FROM ACUTE LYMPHO BLASTIC LEUKEMIA CELLS AND

PRODUCTION OF

ANTI SERA.

AUTHOR(S): SIDDIQUI F A; SAHAI SRIVASTAVA B I

CORPORATE SOURCE: GRACE CANCER DRUG CENT., ROSWELL PARK MEM. INST., BUFFALO,



SOURCE: N.Y. 14263, USA.  
 BIOCHEM BIOPHYS ACTA, (1978) 517 (1 50-157.  
 CODEN: BBACAQ. ISSN: 0006-3002.

FILE SEGMENT: BA; OLD

LANGUAGE: English

AB Terminal deoxyribonucleotidyl transferase (nucleosidetriphosphate:DNA deoxynucleotidylexotransferase, EC 2:7.7.31) was purified from blast cells of a patient with acute lymphoblastic leukemia. The purified enzyme had a specific activity of 31,902 **units/mg** of protein (1 unit equals 1 nmol of Mn<sup>2+</sup> dGTP used on dA12-12 initiator in 1 h) and gave a single protein band on polyacrylamide gel electrophoresis under nondenaturing conditions. In the native state the molecular weight of the enzyme is between 32,000 and 34,000 as determined by glycerol gradient centrifugation and Sephadex G-200 column chromatography. On sodium dodecyl sulfate-polyacrylamide gel electrophoresis it gave 2 bands, 1 corresponding to a subunit of MW 28,000 and the other to 8500.

Antiserum to purified enzyme was prepared in rabbits, using terminal deoxyribonucleotidyl transferase crosslinked to bovine serum albumin and antibody production was detected by immunodiffusion against terminal deoxyribonucleotidyl transferase and neutralization of the enzyme activity. Antibodies to terminal deoxyribonucleotidyl transferase were partially purified by 60% ammonium sulfate precipitation followed by Sephadex G-200 chromatography. This antibody preparation inhibited, in vitro, the activity of the enzyme from the above cells, as well as that from other leukemic cells and calf thymus but it did not inhibit DNA polymerases .alpha., .beta. and .gamma. from any of these sources or **reverse transcriptase** from simian sarcoma virus. Terminal deoxynucleotidyl transferases from calf thymus and human leukemic cells resemble each other in subunit composition and antibody specificity.

L11 ANSWER 11 OF 11 MEDLINE  
 ACCESSION NUMBER: 77022084 MEDLINE  
 DOCUMENT NUMBER: 77022084  
 TITLE: HeLa cell DNA polymerase gamma: further purification and properties of the enzyme.  
 AUTHOR: Knopf K W; Yamada M; Weissbach A  
 SOURCE: BIOCHEMISTRY, (1976 Oct 5) 15 (20) 4540-8.  
 Journal code: A0G. ISSN: 0006-2960.  
 PUB. COUNTRY: United States  
 LANGUAGE: English  
 FILE SEGMENT: Priority Journals  
 ENTRY MONTH: 197702

AB DNA polymerase gamma has been purified over 60 000-fold from HeLa cells which contain no detectable type C viral particles. This purified enzyme shows a specific activity of 25 000 **units/mg** of protein which is comparable to the known specific activity of homogeneous preparations of human alpha and beta polymerases. The isolated enzyme shows apparent molecular weights ranging from 160 000 to 330 000 according to the method of analysis. The enzyme exhibits optimal activity for copying poly(A) in the presence of 50 mM KPO<sub>4</sub> and 130 mM KCl and, under these conditions, copies poly(A) 20 times more rapidly than activated DNA. These assay conditions permit a clear distinction between the gamma-polymerase and DNA polymerase beta which is markedly inhibited by

phosphate at this concentration. A comparison of the copying of activated

DNA, poly(dA) and poly(A) by DNA polymerases alpha, beta, and gamma under

optimal assay conditions for each enzyme is presented. Studies with synthetic and natural nucleic acid templates also show the gamma-polymerase to behave differently than the **reverse transcriptases** of avian myeloblastosis virus or Rauscher leukemia virus.